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(54) Title: USE OF CPG NUCLEIC ACIDS IN PRION-DISEASE

(57) Abstract: Methods are provided which are useful in the treatment of prion diseases and other protein deposit diseases, including, for example, post-exposure prophylaxis against the development of iatrogenic Creutzfeldt-Jakob disease. The methods involve the use of immunostimulatory nucleic acids, including CpG nucleic acids.



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USE OF CpG NUCLEIC ACIDS IN PRION DISEASE

Field of the Invention

The instant invention pertains to methods useful in the treatment of prion diseases, including, for example, post-exposure prophylaxis against the development of iatrogenic Creutzfeldt-Jakob disease. The methods involve the use of immunostimulatory nucleic acids.

Background of the Invention

Prion diseases include a number of fatal, neurodegenerative diseases believed to be caused by aggregates of normal protein that is present in an abnormal conformation. The normal protein, prion protein, is usually present in the cell membrane of many tissues, particularly neuronal tissue. The abnormally conformed prion protein is believed to be directly involved in converting normally conformed prion protein into more of the abnormally conformed prion protein, which then self-assembles into aggregates that are damaging to neuronal tissue anatomy and function.

At least some of the prion diseases are transmissible. However, unlike bacteria, viruses, fungi, parasites, and other replicating pathogens, transmissible prions are simply proteins; they are transmissible without any accompanying nucleic acid. For reasons that are not yet fully understood, the abnormally conformed prion proteins normally do not induce an immune response. Thus, exposure of a healthy individual to abnormally conformed prion protein can initiate a prion disease that can go unchecked by the immune system.

Exposure to abnormally conformed prion protein thus represents a health risk to susceptible individuals. Such individuals include humans and non-humans, principally cows and sheep. Exposure can come about through contact with prion-diseased animal products through ingestion, iatrogenic or work-related exposure, transplantation, and administration of pharmaceutical preparations.

Summary of the Invention

The instant invention is based in part on the unexpected discovery by the inventors that administration of immunostimulatory nucleic acid to a subject that is exposed to abnormally conformed prion protein is an effective treatment of prion disease. The immunostimulatory nucleic acid effectively delayed and even prevented disease in animals

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administered very large doses of prion-diseased brain homogenates that uniformly caused fatal disease in all untreated control animals similarly exposed to prion-diseased brain homogenates.

The immunostimulatory nucleic acids are useful in the treatment of prion diseases, including Creutzfeldt-Jakob disease (CJD), bovine spongiform encephalopathy (BSE), and scrapie. The methods of the invention are also useful for the study of these diseases, for instance, in animal models. The methods will also be useful for developing an understanding of how prion proteins normally fail to elicit an immune response and how an immune response can be elicited and used to treat prion protein disease.

The immunostimulatory nucleic acids are also useful in the treatment of other neurologic diseases involving abnormal protein deposits or aggregates. Such diseases include Alzheimer's disease, which involves deposits of amyloid. The main component of amyloid plaques is amyloid-β peptide (Aβ), a fibrillar 40-42 amino acid peptide that accumulates extracellularly and causes neuronal death.

In one aspect the invention provides a method for treating a prion disease in a subject. The method involves administering to a subject having or at risk of developing a prion disease a CpG nucleic acid in an effective amount to treat the prion disease. In one embodiment the administering follows exposure of the subject to a prion protein that is associated with a prion disease. In one embodiment the prion disease is a transmissible spongiform encephalopathy (TSE). In one embodiment the subject is a human.

In various preferred embodiments, the prion disease is scrapie, BSE, or a form of CJD. The CJD in one embodiment is introgenic CJD (iCJD). In another embodiment the CJD is variant CJD (vCJD). In yet another embodiment the CJD is sporadic CJD.

In one aspect the invention provides a method for inducing an immune response to a prion protein. The method according to this aspect of the invention involves the steps of contacting an antigen-presenting cell (APC) with a prion protein and contacting the APC with a CpG nucleic acid in an effective amount to induce an immune response to the prion protein. In one embodiment the immune response occurs in vivo. In one embodiment the immune response occurs in vitro. In all embodiments according to this aspect of the invention the APC is preferably chosen from a B cell, a dendritic cell, a macrophage, and a monocyte. In one preferred embodiment the APC is a dendritic cell.

Also according to this aspect of the invention, in one embodiment the APC expresses a Toll-like receptor (TLR) that signals in response to the CpG nucleic acid. It has recently been reported that CpG nucleic acid can specifically induce a particular TLR, designated TLR9, to signal. Accordingly, in one embodiment the TLR is TLR9.

In one embodiment the prion protein includes prion protein:scrapie form (PrP^{Sc}). In another embodiment the prion protein includes a fragment of PrP^{Sc} lacking at least the amino terminus of full-length PrP^{Sc}. In yet another embodiment the prion protein includes a derivative of PrP^{Sc} or a derivative of a fragment of PrP^{Sc} lacking at least the amino terminus of full-length PrP^{Sc}.

In one embodiment the prion protein is prion protein:scrapie form (PrPSc). In another embodiment the prion protein is a fragment of PrPSc lacking at least the amino terminus of full-length PrPSc. In yet another embodiment the prion protein is a derivative of PrPSc or a derivative of a fragment of PrPSc lacking at least the amino terminus of full-length PrPSc.

It has been reported that certain CpG nucleic acids are more effective in one species than in another. Accordingly, in preferred embodiments the CpG nucleic acid is optimized for use in a species of the subject.

It has also been reported that CpG nucleic acids appear to fall into different classes based on certain structural features as well as their function. At least three classes are believed to exist, denoted Class A, Class B, and Class C. In one embodiment the CpG nucleic acid is a class B CpG nucleic acid. In one embodiment the CpG nucleic acid is a class A CpG nucleic acid. In one embodiment the CpG nucleic acid is a class C CpG nucleic acid.

These and other features of the invention are described in greater detail in connection with the detailed description of the invention.

Detailed Description of the Invention

A major step in the study of prions and the diseases that they cause was the discovery and purification of a protein designated prion protein (PrP). Bolton et al. (1982) *Science* 218:1309-11; Prusiner SB et al. (1982) *Biochemistry* 21:6942-50; McKinley MP et al. (1983) *Cell* 35:57-62. Complete prion protein-encoding genes have since been cloned, sequenced and expressed in transgenic animals. PrP^C is encoded by a single-copy host gene (Basler K et

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al. (1986) Cell 46:417-28) and is normally found at the outer surface of neurons. Prion diseases are accompanied by the conversion of PrP^C into a modified form called PrP^{Sc}.

The scrapie isoform of the prion protein (PrPSc) is thought necessary for both the transmission and pathogenesis of the transmissible neurodegenerative diseases of animals and humans. See Prusiner SB (1991) *Science* 252:1515-22. The most common prion diseases of animals are scrapie of sheep and goats and bovine spongiform encephalopathy (BSE; "mad cow disease") of cattle. Wilesmith J et al. (1991) *Microbiol Immunol* 172:21-38. Four prion diseases of humans have been identified: (1) kuru, (2) Creutzfeldt-Jakob disease (CJD), (3) Gerstmann-Sträussler-Scheinker (GSS) syndrome, and (4) fatal familial insomnia. Gajdusek DC (1977) *Science* 197:943-60; Medori et al. (1992) *N Engl J Med* 326:444-9.

Most CJD cases are sporadic, but about 10-15% are inherited as autosomal dominant disorders that are caused by mutations in the human PrP gene. Hsiao et al. (1990) Neurology 40:1820-7; Goldfarb et al. (1992) Science 258:806-8. Iatrogenic CJD has been caused by human growth hormone derived from cadaveric pituitaries as well as dura mater grafts. Brown et al. (1992) Lancet 340:24-7. Despite numerous attempts to link CJD to an infectious source such as the consumption of meat from scrapie-infected sheep, none has been identified to date (Harries-Jones et al. (1988) J Neurol Neurosurg Psychiatry 51:1113-9) except in cases of iatrogenically induced disease. On the other hand, kuru, which for many decades devastated the Fore and neighboring tribes of the New Guinea highlands, is believed to have been spread by infection during ritualistic cannibalism.

The major component of purified infectious prions, designated PrP 27-30, is the proteinase K resistant core of a larger native protein PrP^{Sc} which is the disease causing form of the ubiquitous cellular protein PrP^C. PrP^{Sc} is found only in scrapie infected cells, whereas PrP^C is present in both infected and uninfected cells implicating PrP^{Sc} as the major, if not the sole, component of infectious prion particles. Since both PrP^C and PrP^{Sc} are encoded by the same single copy gene, great effort has been directed toward unraveling the mechanism by which PrP^{Sc} is derived from PrP^C. Central to this goal has been the characterization of physical and chemical differences between these two molecules. Properties distinguishing PrP^{Sc} from PrP^C include low solubility (Meyer RK et al. (1986) *Proc Natl Acad Sci USA* 83:2310-4), poor antigenicity (Kascsak RJ et al. (1987) *J Virol* 61:3688-93; Serban D et al. (1990) *Neurology* 40:110-7), protease resistance (Oesch B et al. (1985) *Cell* 40:735-46), and polymerization of PrP 27-30 into rod-shaped aggregates which are very similar, on the

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ultrastructural and histochemical levels, to the PrP amyloid plaques seen in scrapie-diseased brains (Prusiner SB et al. (1983) *Cell* 35(2 Pt 1):349-58). By using proteinase K it is possible to denature PrP^C but not PrP^{Sc}. To date, attempts to identify any post-translational chemical modifications in PrP^C that lead to its conversion to PrP^{Sc} have proven fruitless. Stahl N et al. (1993) Biochemistry 31:5043-53. Consequently, it has been proposed that PrP^C and PrP^{Sc} are in fact conformational isomers of the same molecule.

Conformational description of PrP using conventional techniques has been hindered by problems of solubility and the difficulty in producing sufficient quantities of pure protein. However, PrP^C and PrP^{Sc} are conformationally distinct. Theoretical calculations based upon the amino acid sequences of PrPs from several species have predicted four putative helical motifs in the molecule. Experimental spectroscopic data would indicate that in PrP^C these regions adopt alpha-helical arrangements, with virtually no beta-sheet. Pan et al. (1993) *Proc Natl Acad Sci USA* 90:10962-6). In dramatic contrast, in the same study it was found that PrP^{Sc} and PrP 27-30 possess significant beta-sheet content, which is typical of amyloid proteins. Moreover, studies with extended synthetic peptides, corresponding to PrP amino acid residues 90-145, have demonstrated that these truncated molecules may be converted to either alpha-helical or beta-sheet structures by altering their solution conditions. The transition of PrP^C to PrP^{Sc} requires the adoption of beta-sheet structure by regions that were previously alpha-helical.

In general, scrapie infection fails to produce an immune response, with host organisms being tolerant to PrPSc from the same species. Polyclonal anti-PrP antibodies have been raised in rabbits following immunization with large amounts of Syrian hamster PrP 27-30. Bendheim PE et al. (1985) *Proc Natl Acad Sci USA* 82:997-1001; Bode L et al. (1985) *J Gen Virol* 66:2471-8. Similarly, a handful of anti-PrP monoclonal antibodies have been produced in mice. Kascsak RJ et al. (1987) *J Virol* 61:3688-93; Barry RA et al. (1986) *J Infect Dis* 154:518-21. These antibodies are able to recognize native PrPC and denatured PrPSc from both Syrian hamsters and humans equally well, but do not bind to murine PrP. Unsurprisingly, the epitopes of these antibodies were mapped to regions of sequence containing amino acid differences between Syrian hamster and murine PrP. Rogers et al. (1991) *J Immunol* 147:3568-74.

The DNA sequence of the human, sheep and cow PrP genes have been determined, allowing, in each case, the prediction of the complete amino acid sequence of their respective

PrP proteins. The normal amino acid sequence which occurs in the vast majority of individuals is referred to as the wild-type PrP sequence. This wild-type sequence is subject to certain characteristic polymorphic variations. In the case of human PrP, two polymorphic amino acids occur at residues 129 (Met/Val) and 219 (Glu/Lys). Sheep PrP has two amino acid polymorphisms at residues 136 and 171, while bovine PrP has either five or six repeats of an eight amino acid motif sequence (octarepeats) in the amino terminal region of the mature prion protein. While none of these polymorphisms are of themselves pathogenic, they appear to influence prion diseases. Distinct from these normal variations of the wild-type PrP proteins, certain mutations of the human PrP gene which alter either specific amino acid residues of PrP or the number of octarepeats have been identified which segregate with inherited human prion diseases.

For example, sequences of chicken, bovine, sheep, rat, and mouse PrP genes are disclosed and published within Gabriel JM et al. (1992) *Proc Natl Acad Sci USA* 89:9097-101. A sequence for a PrP gene of Syrian hamster is published in Basler K et al. (1986) *Cell* 46:417-28. A PrP gene of sheep is published by Goldmann W et al. (1990) *Proc Natl Acad Sci USA* 87:2476-80. A gene sequence for bovine PrP is published in Goldmann W et al. (1991) *J Gen Virol* 72:201-4. A sequence for chicken PrP gene is published in Harris DA et al. (1991) *Proc Natl Acad Sci USA* 88:7664-8. A PrP gene sequence for mink is published in Kretzschmar HA et al. (1992) *J Gen Virol* 73:2757-61. A human PrP gene sequence is published in Kretzschmar HA et al. (1986) *DNA* 5:315-24. A PrP gene sequence for mouse is published in Locht C et al. (1986) *Proc Natl Acad Sci USA* 83:6372-6. A PrP gene sequence for sheep is published in Westaway D et al. (1994) *Genes Dev* 8:959-69. These publications are all incorporated herein by reference to disclose and describe the PrP gene and PrP amino acid sequences.

Human PrP cDNA (SEQ ID NO:1; GenBank Accession No. M13899)

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eggegeegeg agetteteet etecteaega eegaggeaga geagteatta teggegaacet
                                                                    60
tggctgctgg atgctggttc tctttgtggc cacatggagt gacctggqcc tctgcaagaa
                                                                   120
gcgcccgaag cctggaggat ggaacactgg gggcagccga tacccggggc agggcagccc
                                                                   180
tggaggcaac cgctacccac ctcagggcgg tggtggctgg gggcagcctc atggtggtgg
                                                                   240
ctgggggcag cctcatggtg gtggctgggg gcagccccat ggtggtggct ggggacagcc
                                                                   300
tcatggtggt ggctggggtc aaggaggtgg cacccacagt cagtggaaca agccgagtaa
                                                                   360
gccaaaaacc aacatgaagc acatggctgg tgctgcagca gctggggcag tggtggggg
                                                                   420
ccttggcggc tacatgctgg gaagtgccat gagcaggccc atcatacatt tcggcagtga
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Mink PrP genomic DNA (SEQ ID NO:2; GenBank Accession No. S46825)

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  gaaatacgca tgtgcttata ttttttatat ttgtaacttt gcatgtactt gttttgtgtt 3960
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                                                                     120
  tacccaggac agggcagtcc tggaggcaac cgttatccac ctcagggagg gggtggctgg 180
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  accaccacca ccaaggggga gaacttcacc gaaactgaca tcaagatgat ggagcgagtg 660
  gtggagcaaa tgtgcattac ccagtaccag agagaatccc aggcttatta ccaacgaggg 720
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                                                                     780
                                                                      795
  ctcatagtag gatag
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- 10 -

Chicken PrP cDNA (SEQ ID NO:6; GenBank Accession No. M61145) quattecete ggeagecage tectecetet egetatttat teetttetee ecceetaeg 60 ctggatctgg atcatctcaa gccgagcggt gacggcttct tggatcgctc atacataaat 120 atctqtqaqt cagagqaagc aaccaccgac cccaagacct caccccgagc catggctagg 180 ctecteacea cetgetgeet getggeeetg etgetegeeg cetgeacega egtegeeete 240 tccaaqaagg gcaaaggcaa acccagtggt gggggttggg gcgccgggag ccatcgccag 300 cccagctacc cccgccagcc gggctaccct cataacccag ggtaccccca taacccaggg 360 tacceccaca accetggeta tececataae ceeggetace eccagaacee tggetacece 420 cataacccag gttacccagg ctggggtcaa ggctacaacc catccagcgg aggaagttac 480 cacaaccaga agccatggaa accccccaaa accaacttca agcacgtggc gggggcagca 540 geggeggtg etgtggtggg gggettgggg ggetaegeea tggggegegt tatgteaggg 600 atgaactacc acttcgatag acccgatgag taccgatggt ggagtgagaa ctcggcgcgt 660 tateceaace gggtttaeta eegggattae ageageeeeg tgecacagga egtettegtg 720 gccgattgct ttaacatcac agtgactgag tacagcattg gccctgctgc caagaagaac acctccgagg ctgtggcggc agcaaaccaa acggaggtgg agatggagaa caaagtggtg 840 acqaaqqtqa teeqeqaqat qtqeqtqeaq caqtaceqeg agtaceqeet ggeeteggge atccagetge accetgetga cacetggete geegteetee teeteeteet caceaccett 960 tttgccatgc actgatggga tgccgtgccc cggccctgtg gcagtgagat gacatcgtgt 1020 eccegtgeec acceatgggg tgtteettgt cetegetttt gtecatettt ggtgaagatg 1080 tecceeeget geeteeege aggetetgat ttgggeaaat gggaggggat tttgteetgt 1140 cctggtcgtg gcaggacggc tgctggtggt ggagtgggat gcccaaaaaa tggccttcac 1200 cactteetee teetetteet ttetggggeg gagatatggg etegteeage cettattgte 1260 cctgcaagag cgtatctgaa aatcctcttt gctaacaagc agggttttac ctaatctgct 1320 tagccccagt gacagcagag cgcctttccc cagggcacac caaccccaag ctgaggtgct 1380 tggcagccac acgtcccatg gaggctgatg ggttttgggg cgtcccaagc aacaccctgg 1440 gctactgagg tgcaattgta gctctttaat ctgccaatcc caaccctacc gtgtagatag 1500 gaactgeetg etetgeattt tgeatgetge aaacacetee tgeegeageg eececaaaat 1560 agagtgattt gggaatagtg aggetgaage cacageaget tgggattggg etcatcatat 1620 caatccatga tgctttgctt ccagctgagc ctcactgccc ttttatagcc tgcccagagg 1680 aaqqqaqcqc tqctaaatgc ccaaaaaggt aacactgagc aaaagcttat ttcaatgtat 1740 qataqagaac qaqtqcatct cgcacagatc agccatggga gcatcgtttg ccatcagccc 1800 caaaacccaa aggatgctaa aatgcagcca aaggggaatc aagcacgcag ggaaggactt 1860 gaatcagctc aactggattg aaatggcaaa aggcatgagt agaacgaacg gcaaggggat 1920 gctggagatc cacctcctgt gagcaaattg ttcgatgcag ccaatggaac tattgcttct 1980 tgtgcttcag ttgctgctga tgtgtacata ggctgtagca tatgtaaagt tacacgtgtc 2040 aagctgctcg caccgcgtag agctaatatg tatcatgtat gtgggcactg aatgccaccg 2100 ttggccatac ccaaccgtcc taaacgattt tcacgtcgct gtaacttaag tggagataca 2160 ctttcagtat attcagcaaa aggaattc 2188 Mouse PrP cDNA (SEO ID NO:7; GenBank Accession No. M13685) aatteettea qaactqaace attteaaceq aqetqaaqea ttetqeette etaqtqqtae caqtccaatt taggagagcc aagcagacta tcagtcatca tggcgaacct tggctactgg 120 ctqctqqccc tctttqtqac tatqtqqact qatqtcqqcc tctqcaaaaa gcqqccaaaq 180 cctggagggt ggaacaccgg tggaagccgg tatcccgggc agggaagccc tggaggcaac 240 300 cgttacccac ctcagggtgg cacctggggg cagccccacg gtggtggctg gggacaaccc catgggggca gctggggaca acctcatggt ggtagttggg gtcagcccca tggcggtgga 360 tggggccaag gagggggtac ccataatcag tggaacaagc ccagcaaacc aaaaaccaac 420 ctcaagcatg tggcagggc tgcggcagct ggggcagtag tggggggcct tggtggctac 480 atgctgggga gcgccgtgag caggcccatg atccattttg gcaacgactg ggaggaccgc 540 tactaccgtg aaaacatgta ccgctaccct aaccaagtgt actacaggcc agtggatcag 600 tacagcaacc agaacaactt cgtgcacgac tgcgtcaata tcaccatcaa gcagcacacg 660

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agaagateca geageacegt gettttetee teceeteetg teatecteet cateteette

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780

840

900

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  tqaatctqca tgtacttcac gttttctata tttgtaactt tgcatgtatt ttgttttgtc 2040
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Sheep PrP cDNA (SEQ ID NO:8; GenBank Accession No. X79912)
  qcaqaqaaqt catcatqqtq aaaagccaca taggcagttg gatcctggtt ctctttgtgg
                                                                     60
  ccatgtggag tgacgtgggc ctctgcaaga agcgaccaaa acctggcgga ggatggaaca 120
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  tectettte tteccetect gtgatectee teatetett ceteatttt eteatagtag 780
                                                                    803
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Rat PrP cDNA (SEQ ID NO:9; GenBank Accession No. NM 012631)
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  catggtggtg gctggggaca acctcatggt ggtggctggg gacaacctca tggtggtggc
                                                                    240
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                                                                    300
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                                                                    360
                                                                    420
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  tttggcaacq actgggagga ccgctactac cgagaaaaca tgtaccgtta ccctaaccaa
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                                                                    720
                                                                    765
  gtgatectee teateteett ceteatette etgategtgg gatga
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As used herein, "prion disease" refers to any disease or condition in a subject, the pathogenesis of which involves a prion protein other than PrP^C of the species of the subject.

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A prion disease will typically but not necessarily be a transmissible spongiform encephalopathy.

As used herein, "transmissible spongiform encephalopathy" and, equivalently, "(TSE)" shall mean any prion disease that is associated with spongiform encephalopathy and is communicable from one individual to another. As prion diseases can include entities other than TSE, this term refers to at least a subset of all prion diseases. At present TSE includes Creutzfeldt-Jakob disease, kuru, Gerstmann-Sträussler-Scheinker syndrome, fatal familial insomnia, bovine spongiform encephalopathy, and scrapie.

As used herein, "Creutzfeldt-Jakob disease" and, equivalently, "(CJD)" refers to the TSE that naturally occurs in humans. CJD includes sporadic, genetic (familial), and infectious (i.e., variant and iatrogenic) forms. CJD is a well described entity in the medical literature, and until now has been widely believed to be a uniformly fatal neurodegenerative disease for which there is no effective form of treatment.

As used herein, "variant Creutzfeldt-Jakob disease" and, equivalently, "(vCJD)", also referred to in the literature as new variant Creutzfeldt-Jakob disease (nvCJD), refers to CJD attributable to the BSE prion. It can be distinguished from sporadic CJD not only by the prion involved but also by certain clinical and preclinical features. See Aguzzi A (2000) Haematologica 85:3-10; Hill AF et al. (1997) Nature 389:448-50; Bruce ME et al. (1997) Nature 389:498-501; Will R et al. (1996) Lancet 347:921-5.

As used herein, "iatrogenic Creutzfeldt-Jakob disease" and, equivalently, "(iCJD)" refers to any form of CJD that is attributable to work- or treatment-related exposure to prion protein that is associated with CJD.

As used herein, "bovine spongiform encephalopathy" and, equivalently, "(BSE)" shall refer to the TSE that occurs naturally in cows and cattle.

As used herein, "scrapie" refers to the TSE that occurs naturally in sheep and goats, as well as to experimental models of scrapie. Scrapie in sheep has been recognized and described in the literature for over 300 years. For a review, see O'Rourke KI (2001) Vet Clin North Am Food Anim Pract 17:283-300.

As used herein, a "prion protein that is associated with a prion disease" refers to any prion protein involved in the pathogenesis of a prion disease. A prion protein that is associated with a prion disease can be a prion found in nature. Alternatively, prion protein that is associated with a prion disease can be a prion protein made de novo or modified from

its natural form through human activity, e.g., by in vitro synthesis, chemical synthesis, chemical derivativization, or genetic alteration. Chemical alteration includes, without limitation, altered glycosylation. In a preferred embodiment, a prion protein that is associated with a prion disease is a prion protein found in nature, e.g., PrPSc. In one embodiment, a prion protein that is associated with a prion disease is a truncated or genetically modified form of a prion protein found in nature. A genetically modified form of a prion protein found in nature includes a fusion protein involving at least a substantial portion of a prion protein as one component, a prion protein that differs from a prion protein found in nature by one or more conservative amino acid substitutions, and allelic variants of the prion protein found in nature.

Naturally occurring residues can be divided into the following classes based on common side chain properties: (1) hydrophobic: norleucine, Met, Ala, Val, Leu, Ile; (2) neutral hydrophilic: Cys, Ser, Thr; (3) acidic: Asp, Glu; (4) basic: Asp, Gln, His, Lys, Arg; (5) residues that influence chain orientation: Gly, Pro; and (6) aromatic: Trp, Tyr, Phe. Thus, for example, conservative amino acid substitutions can involve the exchange of a member from one of these classes for another member from the same class. Non-conservative amino acid substitutions can involve the exchange of a member of one of these classes for a member from another class.

A conservative amino acid substitution can involve a substitution of a native amino acid residue with another residue such that there is little or no effect on the polarity or charge of the amino acid residue at that position. Conservative amino acid substitutions also encompass non-naturally occurring amino acid residues that are typically incorporated by chemical peptide synthesis rather than by synthesis in biological systems. These include peptidomimetics, and other reversed or inverted forms of amino acid moieties.

As used herein, "prion protein:scrapie form (PrPSc)" refers to any of a number of naturally occurring, species-specific, proteinase K-resistant forms of prion protein associated with prion disease. Specific examples include, but are not limited to, the following: human PrPSc having an amino acid sequence provided by SEQ ID NO:10; bovine PrPSc having an amino acid sequence provided by SEQ ID NO:11; bovine PrPSc having an amino acid sequence provided by SEQ ID NO:12; ovine PrPSc having an amino acid sequence provided by SEQ ID NO:13; ovine PrPSc having an amino acid sequence provided by SEQ ID NO:14; and murine PrPSc having an amino acid sequence provided by SEQ ID NO:15.

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H	uman PrP ^{sc} (S	EQ ID NO:10	; GenBank A	ccession No. A	AAE81600)		
	MANLGCWMLV	LFVATWSDLG	LCKKRPKPGG	WNTGGSRYPG	QGSPGGNRYP	PQGGGGWGQP	60
	HGGGWGQPHG	GGWGQPHGGG	WGQPHGGGWG	QGGGTHSQWN	KPSKPKTNMK	HMAGAAAAGA	120
	VVGGLGGYML	GSAMSRPIIH	FGSDYEDRYY	RENMHRYPNQ	VYYRPMDEYS	NQNNFVHDCV	180
	NITIKQHTVT	TTTKGENFTE	TDVKMMERVV	EQMCITQYER	ESQAYYQRGS	SMVLFSSPPV	240
	ILLISFLIFL	IVG					253
В	ovine PrP ^{Sc} (S	EQ ID NO:11	; GenGank A	ccession No. A	AAE81601)		
	MVKSHIGSWI	LVLFVAMWSD	VGLCKKRPKP	GGWNTGGSRY	PGQGSPGGNR	YPPQGGGGWG	60
	QPHGGGWGQP	HGGGWGQPHG	GGWGQPHGGG	WGQPHGGGGW	GQGGTHGQWN	KPSKPKTNMK	120
	HVAGAAAAGA	VVGGLGGYML	GSAMSRPLIH	FGSDYEDRYY	RENMHRYPNQ	VYYRPVDQYS	180
	NQNNFVHDCV	NITVKEHTVT	TTTKGENFTE	TDIKMMERVV	EQMCVTQYQK	ESQAYYDQGA	240
	SVILFSSPPV	ILLISFLIFL	IVG				263
В	ovine PrP ^{Sc} (S	EQ ID NO:12	; GenGank A	ccession No. (CAA39368)		
	MVKSHIGSWI	LVLFVAMWSD	VGLCKKRPKP	GGGWNTGGSR	YPGQGSPGGN	RYPPQGGGGW	60
	GQPHGGGWGQ	PHGGGWGQPH	GGGWGQPHGG	GWGQPHGGGG	WGQGGTHGQW	NKPSKPKTNM	120
	KHVAGAAAAG	AVVGGLGGYM	LGSAMSRPLI	HFGSDYEDRY	YRENMHRYPN	QVYYRPVDQY	180
	SNQNNFVHDC	VNITVKEHTV	TTTTKGENFT	ETDIKMMERV	VEQMCITQYQ	RESQAYYQRG	240
	ASVILFSSPP	VILLISFLIF	LIVG				26
O	vine PrP ^{Sc} (SE	EQ ID NO:13;	GenBank Acc	cession No. A	AE81602)		
	MVKSHIGSWI	LVLFVAMWSD	VGLCKKRPKP	GGWNTGGSRY	PGQGSPGGNR	YPPQGGGGWG	60
	QPHGGGWGQP	HGGGWGQPHG	GSWGQPHGGG	GWGQGGSHSQ	WNKPSKPKTN	MKHVAGAAAA	120
	GAVVGGLGGY	MLGSAMSRPL	IHFGNDYEDR	YYRENMYRYP	NQVYYRPVDQ	YSNQNNFVHD	180
	CVNITVKQHT	VTTTTKGENF	TETDIKIMER	VVEQMCITQY	QRESQAYYQR	GASVILFSSP	240
	PVILLISFLI	FLIVG					255
O	vine PrP ^{Sc} (SE	EQ ID NO:14;	GenBank Ac	cession No. C	AA56283)		
	MVKSHIGSWI	LVLFVAMWSD	VGLCKKRPKP	GGGWNTGGSR	YPGQGSPGGN	RYPPQGGGGW	60
	GQPHGGGWGQ	PHGGGWGQPH	GGGWGQPHGG	GGWGQGGSHS	QWNKPSKPKT	NMKHVAGAAA	120
	AGAVVGGLGG	YMLGSAMSRP	LIHFGNDYED	RYYRENMYRY	PNQVYYRPVD	QYSNQNNFVH	180
	DCVNITVKQH	TVTTTTKGEN	FTETDIKIME	RVVEQMCITQ	YQRESQAYYQ	RGASVILFSS	240
	PPVILLISFL	IFLIVG				•	256

Murine PrPSc (SEQ ID NO:15; GenBank Accession No. AAE81599)

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MANLGYWLLA	LFVTMWTDVG	LCKKRPKPGG	WNTGGSRYPG	QGSPGGNRYP	PQGGTWGQPH	60
GGGWGQPHGG	SWGQPHGGSW	GQPHGGGWGQ	GGGTHNQWNK	PSKPKTNLKH	VAGAAAAGAV	120
VGGLGGYMLG	SAMSRPMIHF	GNDWEDRYYR	ENMYRYPNQV	YYRPVDQYSN	QNNFVHDCVN	180
ITIKQHTVTT	TTKGENFTET	DVKMMERVVE	QMCVTQYQKE	SQAYYDGRRS	SSTVLFSSPP	240
VILLISELIE	LIVG					254

As used herein, "full-length PrP^{Sc}" refers to a form of PrP^{Sc} that includes all its amino acids as it occurs in nature. It is to be distinguished, for example, from a truncated form of PrP^{Sc}, described elsewhere herein. A full-length PrP^{Sc} can, however, be incorporated into a PrP^{Sc} conjugate or PrP^{Sc} fusion protein.

As used herein, a "derivative of PrPSc" refers to a chemical or genetic derivative of a naturally occurring form of PrPSc, including PrPSc with non-native glycosylation, covalent or non-covalent conjugates formed between PrPSc and another compound, PrPSc fusion proteins, and any combination thereof. A "derivative of a fragment of PrPSc lacking at least the amino terminus of full-length PrPSc" refers to a chemical or genetic derivative of an N-terminally truncated form of PrPSc, including such truncated forms of PrPSc: (i) with non-native glycosylation, (ii) as part of a covalent or non-covalent conjugate formed with another compound, (iii) as part of a fusion protein, or (iv) any combination thereof. In preferred embodiments the fragment of PrPSc lacking at least the amino terminus of full-length PrPSc refers to a fragment lacking one or more, up to and including all, of the octarepeats (e.g., GGGWGQPH (SEQ ID NO:16) and GGSWGQPH (SEQ ID NO:17)). See Flechsig E et al. (2000) Neuron 27:399-408.

A truncated form of a prion protein found in nature is identical in primary sequence to the prion protein found in nature except for the absence of one or more amino acid residues from the N-terminal end, the C-terminal end, or both the N-terminal and the C-terminal ends. Truncated forms can also include deletion mutants, in which an internal sequence is omitted without changing either the N-terminal end or the C-terminal end. In a preferred embodiment, a truncated prion protein lacks one or more, up to and including all, N-terminal octarepeats (e.g., GGGWGQPH and GGSWGQPH). See Flechsig E et al. (2000) Neuron 27:399-408.

As used herein, a "fragment of PrPSc lacking at least the amino terminus of full-length PrPSc" shall refer to a truncated form of full-length PrPSc lacking one or more N-terminal amino acids normally present in full-length PrPSc. In a preferred embodiment, the fragment

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lacks one or more, up to and including all, of the octarepeats (e.g., GGGWGQPH and GGSWGQPH). See Flechsig E et al. (2000) Neuron 27:399-408.

The methods of the instant invention employ immunostimulatory nucleic acids. In the preferred embodiment, the immunostimulatory nucleic acid is a CpG nucleic acid. The terms "nucleic acid" and "oligonucleotide" are used interchangeably to mean multiple nucleotides (i.e., molecules comprising a sugar (e.g., ribose or deoxyribose) linked to a phosphate group and to an exchangeable organic base, which is either a substituted pyrimidine (e.g., cytosine (C), thymidine (T) or uracil (U)) or a substituted purine (e.g., adenine (A) or guanine (G)). As used herein, the terms "nucleic acid" and "oligonucleotide" refer to oligoribonucleotides as well as oligodeoxyribonucleotides. The terms "nucleic acid" and "oligonucleotide" shall also include polynucleosides (i.e., a polynucleotide minus the phosphate) and any other organic base containing polymer. Nucleic acid molecules can be obtained from existing nucleic acid sources (e.g., genomic or cDNA), but are preferably synthetic (e.g., produced by nucleic acid synthesis).

The terms "nucleic acid" and "oligonucleotide" also encompass nucleic acids or oligonucleotides with substitutions or modifications, such as in the bases and/or sugars. For example, they include nucleic acids having backbone sugars that are covalently attached to low molecular weight organic groups other than a hydroxyl group at the 2' position and other than a phosphate group at the 5' position. Thus modified nucleic acids may include a 2'-O-alkylated ribose group. In addition, modified nucleic acids may include sugars such as arabinose instead of ribose. Thus the nucleic acids may be heterogeneous in backbone composition thereby containing any possible combination of polymer units linked together such as peptide-nucleic acids (which have an amino acid backbone with nucleic acid bases).

Nucleic acids also include substituted purines and pyrimidines such as C-5 propyne modified bases. Wagner RW et al. (1996) Nat Biotechnol 14:840-4. Purines and pyrimidines include but are not limited to adenine, cytosine, guanine, thymidine, 5-methylcytosine, 2-aminopurine, 2-amino-6-chloropurine, 2,6-diaminopurine, hypoxanthine, and other naturally and non-naturally occurring nucleobases, substituted and unsubstituted aromatic moieties. Other such modifications are well known to those of skill in the art.

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The immunostimulatory oligonucleotides of the instant invention can encompass various chemical modifications and substitutions, in comparison to natural RNA and DNA, involving a phosphodiester internucleoside bridge, a β -D-ribose unit and/or a natural

nucleoside base (adenine, guanine, cytosine, thymine, uracil). Examples of chemical modifications are known to the skilled person and are described, for example, in Uhlmann E et al. (1990) Chem Rev 90:543; "Protocols for Oligonucleotides and Analogs" Synthesis and Properties & Synthesis and Analytical Techniques, S. Agrawal, Ed, Humana Press, Totowa, USA 1993; Crooke ST et al. (1996) Annu Rev Pharmacol Toxicol 36:107-129; and Hunziker J et al. (1995) Mod Synth Methods 7:331-417. An oligonucleotide according to the invention can have one or more modifications, wherein each modification is located at a particular phosphodiester internucleoside bridge and/or at a particular β-D-ribose unit and/or at a particular natural nucleoside base position in comparison to an oligonucleotide of the same sequence which is composed of natural DNA or RNA.

For example, the invention relates to an oligonucleotide which comprises one or more modifications and wherein each modification is independently selected from:

- a) the replacement of a phosphodiester internucleoside bridge located at the 3' and/or the 5' end of a nucleoside by a modified internucleoside bridge,
- b) the replacement of phosphodiester bridge located at the 3' and/or the 5' end of a nucleoside by a dephospho bridge,
- c) the replacement of a sugar phosphate unit from the sugar phosphate backbone by another unit,
- d) the replacement of a β -D-ribose unit by a modified sugar unit, and
- e) the replacement of a natural nucleoside base by a modified nucleoside base.

 More detailed examples for the chemical modification of an oligonucleotide are as follows.

A phosphodiester internucleoside bridge located at the 3' and/or the 5' end of a nucleoside can be replaced by a modified internucleoside bridge, wherein the modified internucleoside bridge is for example selected from phosphorothioate, phosphorodithioate, NR 1 R 2 -phosphoramidate, boranophosphate, α -hydroxybenzyl phosphonate, phosphate-(C_1 - C_{21})-O-alkyl ester, phosphate-[(C_6 - C_{12})aryl-(C_1 - C_{21})-O-alkyl]ester, (C_1 - C_8)alkyl-phosphonate and/or (C_6 - C_{12})-arylphosphonate bridges, (C_7 - C_{12})- α -hydroxymethyl-aryl (e.g., disclosed in WO 95/01363), wherein (C_6 - C_{12})aryl, (C_6 - C_{20})aryl and (C_6 - C_{14})aryl are optionally substituted by halogen, alkyl, alkoxy, nitro, cyano, and where R^1 and R^2 are, independently of each other, hydrogen, (C_1 - C_{18})-alkyl, (C_6 - C_{20})-aryl, (C_6 - C_{14})-aryl-(C_1 - C_8)-alkyl, preferably hydrogen, (C_1 - C_8)-alkyl, preferably (C_1 - C_4)-alkyl and/or methoxyethyl, or R^1 and R^2 form, together with

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the nitrogen atom carrying them, a 5-6-membered heterocyclic ring which can additionally contain a further heteroatom from the group O, S and N.

The replacement of a phosphodiester bridge located at the 3' and/or the 5' end of a nucleoside by a dephospho bridge (dephospho bridges are described, for example, in Uhlmann E and Peyman A in "Methods in Molecular Biology", Vol. 20, "Protocols for Oligonucleotides and Analogs", S. Agrawal, Ed., Humana Press, Totowa 1993, Chapter 16, pp. 355ff), wherein a dephospho bridge is for example selected from the dephospho bridges formacetal, 3'-thioformacetal, methylhydroxylamine, oxime, methylenedimethyl-hydrazo, dimethylenesulfone and/or silyl groups.

A sugar phosphate unit (i.e., a β-D-ribose and phosphodiester internucleoside bridge together forming a sugar phosphate unit) from the sugar phosphate backbone (i.e., a sugar phosphate backbone is composed of sugar phosphate units) can be replaced by another unit, wherein the other unit is for example suitable to build up a "morpholino-derivative" oligomer (as described, for example, in Stirchak EP et al. (1989) *Nucleic Acids Res* 17:6129-41), that is, e.g., the replacement by a morpholino-derivative unit; or to build up a polyamide nucleic acid ("PNA"; as described for example, in Nielsen PE et al. (1994) *Bioconjug Chem* 5:3-7), that is, e.g., the replacement by a PNA backbone unit, e.g., by 2-aminoethylglycine.

A β-ribose unit or a β-D-2'-deoxyribose unit can be replaced by a modified sugar unit, wherein the modified sugar unit is for example selected from β-D-ribose, α-D-2'-deoxyribose, L-2'-deoxyribose, 2'-F-2'-deoxyribose, 2'-O-(C₁-C₆)alkyl-ribose, preferably 2'-O-(C₁-C₆)alkyl-ribose is 2'-O-methylribose, 2'-O-(C₂-C₆)alkenyl-ribose, 2'-[O-(C₁-C₆)alkyl-O-(C₁-C₆)alkyl]-ribose, 2'-NH₂-2'-deoxyribose, β-D-xylo-furanose, α-arabinofuranose, 2,4-dideoxy-β-D-erythro-hexo-pyranose, and carbocyclic (described, for example, in Froehler J (1992) *Am Chem Soc* 114:8320) and/or open-chain sugar analogs (described, for example, in Vandendriessche et al. (1993) *Tetrahedron* 49:7223) and/or bicyclosugar analogs (described, for example, in Tarkov M et al. (1993) *Helv Chim Acta* 76:481).

A natural nucleoside base can be replaced by a modified nucleoside base, wherein the modified nucleoside base is for example selected from hypoxanthine, uracil, dihydrouracil, pseudouracil, 2-thiouracil, 4-thiouracil, 5-aminouracil, 5-(C₁-C₆)-alkyluracil, 5-(C₂-C₆)-alkynyluracil, 5-(hydroxymethyl)uracil, 5-chlorouracil, 5-fluorouracil, 5-bromouracil, 5-hydroxycytosine, 5-(C₁-C₆)-alkylcytosine, 5-(C₂-C₆)-alkynylcytosine, 5-chlorocytosine, 5-fluorocytosine,

5-bromocytosine, N²-dimethylguanosine, 2,4-diamino-purine, 8-azapurine, a substituted 7-deazapurine, preferably 7-deaza-7-substituted and/or 7-deaza-8-substituted purine or other modifications of a natural nucleoside bases. This list is meant to be exemplary and is not to be interpreted to be limiting.

The oligonucleotides of the present invention are nucleic acids that contain specific sequences found to elicit an immune response. These specific sequences that elicit an immune response are referred to as "immunostimulatory motifs", and the oligonucleotides that contain immunostimulatory motifs are referred to as "immunostimulatory nucleic acid molecules" and, equivalently, "immunostimulatory nucleic acids" or "immunostimulatory oligonucleotides". The immunostimulatory oligonucleotides of the invention thus include at least one immunostimulatory motif.

In one embodiment of the invention the immunostimulatory oligonucleotides include immunostimulatory motifs which are "CpG dinucleotides". A CpG dinucleotide can be methylated or unmethylated. An immunostimulatory nucleic acid containing at least one unmethylated CpG dinucleotide is a nucleic acid molecule which contains an unmethylated cytosine-guanine dinucleotide sequence (i.e., an unmethylated 5' cytosine followed by 3' guanosine and linked by a phosphate bond) and which activates the immune system; such an immunostimulatory nucleic acid is a CpG nucleic acid. CpG nucleic acids have been described in a number of issued patents, published patent applications, and other publications, including U.S. Patent Nos. 6,194,388; 6,207,646; 6,214,806; 6,218,371; 6,239,116; and 6,339,068.

An immunostimulatory nucleic acid containing at least one methylated CpG dinucleotide is a nucleic acid which contains a methylated cytosine-guanine dinucleotide sequence (i.e., a methylated 5' cytosine followed by a 3' guanosine and linked by a phosphate bond) and which activates the immune system. In other embodiments the immunostimulatory oligonucleotides are free of CpG dinucleotides. These oligonucleotides which are free of CpG dinucleotides are referred to as non-CpG oligonucleotides, and they have non-CpG immunostimulatory motifs. The invention, therefore, also encompasses nucleic acids with other types of immunostimulatory motifs, which can be methylated or unmethylated. The immunostimulatory oligonucleotides of the invention, further, can include any combination of methylated and unmethylated CpG and non-CpG

immunostimulatory motifs. In some embodiments the immunostimulatory oligonucleotide is not an antisense oligonucleotide.

As used herein, a "Toll-like receptor (TLR) that signals in response to the CpG nucleic acid" refers to any TLR that engages or initiates an intracellular signaling pathway associated with the development of an immune response, as a result of contacting the TLR with CpG nucleic acid. The pathway typically involves the adaptor protein MyD88 and subsequent downstream molecules including TRAF, IRAK, Jun, Erk, p38 MAPK, and NF-kB. The TLRs are a family of at least ten highly conserved receptors that share as a common feature a cytoplasmic Toll homology IL-1 receptor (TIR) domain believed to be involved in such signaling. TLR9 is reported to be the natural receptor for CpG nucleic acid.

As to CpG nucleic acids, it has recently been described that there are different classes of CpG nucleic acids. One class is potent for activating B cells but is relatively weak in inducing IFN-α and NK cell activation; this class has been termed the B class. The B class CpG nucleic acids typically are fully stabilized and include an unmethylated CpG dinucleotide within certain preferred base contexts. See, e.g., U.S. Patent Nos. 6,194,388; 6,207,646; 6,214,806; 6,218,371; 6,239,116; and 6,339,068. Another class is potent for inducing IFN-α and NK cell activation but is relatively weak at stimulating B cells; this class has been termed the A class. The A class CpG nucleic acids typically have stabilized poly-G sequences at 5' and 3' ends and a palindromic phosphodiester CpG dinucleotide-containing sequence of at least 6 nucleotides. See, for example, published patent application PCT/US00/26527 (WO 01/22990). Yet another class of CpG nucleic acids activates B cells and NK cells and induces IFN-α; this class has been termed the C class. The C class CpG nucleic acids typically are fully stabilized, include a B class-type sequence and a GC-rich palindrome or near-palindrome. This class has been described in published patent application PCT/US02/26468 (WO 03/015711), the entire content of which is incorporated herein by reference.

Immunostimulatory oligonucleotides are effective in vertebrates. Different immunostimulatory oligonucleotides can cause optimal immune stimulation depending on the type of subject and the sequence of the immunostimulatory oligonucleotide. Many vertebrates have been found according to the invention to be responsive to the same class of immunostimulatory oligonucleotides, sometimes referred to as human specific immunostimulatory oligonucleotides. Rodents, however, respond to different nucleic acids.

Immunostimulatory oligonucleotides causing optimal stimulation in humans may not generally cause optimal stimulation in a mouse and vice versa. An immunostimulatory oligonucleotide causing optimal stimulation in humans often does, however, cause optimal stimulation in other animals such as cow, horses, sheep, etc. For example, within Class B CpG ODN, preferred immunostimulatory sequences have been identified for use in mice (ODN 1826, 5'- TCCATGACGTTCCTGACGTT -3', SEQ ID NO:18) and for use in humans (ODN 2006, 5'- TCGTCGTTTTGTCGTTTTGTCGTT -3', SEQ ID NO:19). One of skill in the art can identify the optimal immunostimulatory nucleic acid sequences useful for a particular species of interest using routine assays described herein and/or known in the art, using the guidance supplied herein.

As used herein, the term "treat" as used in reference to a disease or condition shall mean to intervene in such disease or condition so as to prevent or slow the development of, slow the progression of, halt the progression of, or eliminate the disease or condition. Thus the phrase "to treat the prion disease" as used herein means to prevent or slow the development of, slow the progression of, halt the progression of, or eliminate the prion disease.

As used herein, a "subject" refers to a human or non-human vertebrate. Preferred non-human vertebrates include feed livestock susceptible to TSE, including cows and cattle, sheep, goats, and pigs. Non-human subjects also specifically include non-human primates as well as rodents. Non-human subjects also include, without limitation, chickens, horses, dogs, cats, guinea pigs, hamsters, mink, and rabbits.

As used herein, a "subject having a prion disease" is a subject known or diagnosed to have a prion disease as disclosed herein. Generally a subject having a prion disease will have some objective manifestation of the prion disease, such as a sign, symptom, or result of a suitable diagnostic test that indicates the presence of a prion disease. In the transmissible spongiform encephalopathies, such objective manifestations can include dementia, ataxia, myoclonus, tremor, presence of protease-resistant prion protein in brain extract, and typical or characteristic abnormalities on brain CT, brain MRI, and/or EEG. This list is not meant to be limiting in any way, and those of skill in the art will recognize what criteria are suitable for making a diagnosis of prion disease in a given species in question. A subject having a prion disease shall also include any subject having a test result which specifically indicates the presence in that subject of any amount of prion protein that is associated with a prion

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disease, since it is believed that prion protein that is associated with a prion disease is not present in a subject without a prion disease.

A subject having a prion disease can but need not necessarily have an identifiable risk factor for having a prion disease. An identifiable risk factor for having a prion disease can include a family history of prion disease, a history of consuming known or suspected prion-diseased tissue, or a history of exposure to a prion protein that is associated with a prion disease or to a product derived from a known or suspected prion-diseased tissue (e.g., through administration of pituitary extract).

As used herein, a "subject at risk of developing a prion disease" is a subject with a known or suspected exposure to prion-diseased tissue, a known or suspected exposure to prion protein that is associated with a prion disease, or a known or suspected predisposition to develop a prion disease (e.g., family history of prion disease). In one embodiment the subject at risk of developing a prion disease is a subject residing in or traveling to an area in which TSE is endemic. In one embodiment the subject at risk of developing a prion disease is a subject residing in or traveling to an area in which food or water contains or is likely to contain prion protein that is associated with prion disease. In one embodiment, a subject at risk of developing a prion disease is a subject with a known or suspected iatrogenic exposure to prion-diseased tissue, e.g., neurosurgeons, neuropathologists, pathologists, nurses, morticians, histology technicians and laboratory workers at special risk of contracting iCJD. In one embodiment, a subject at risk of developing a prion disease is a subject with a known or suspected iatrogenic exposure to prion-diseased tissue through receiving a tissue or organ allograft from a subject having a prion disease. Such tissues can include, without limitation, corneas and dural grafts.

As used herein, an "effective amount" of a substance generally refers to that amount of the substance that is sufficient to bring about a desired effect. With reference to CpG nucleic acid, an "effective amount to induce an immune response to the prion protein" shall refer to that amount of CpG nucleic acid that is sufficient to induce an immune response to a particular prion protein. The immune response can occur in vitro, in vivo, ex vivo, and any combination thereof. An immune response to a prion protein can be measured using any suitable means to determine that an immune response occurs in association with exposure of an immune cell to the prion protein. The immune response can be antigen-specific, including any of the following: production of prion protein-specific antibody, proliferation of prion

protein-specific lymphocytes, and cell-mediated immunity against cells expressing prion protein. The immune response can alternatively or additionally be antigen-nonspecific, including any of the following: induction of a Toll-like receptor signaling pathway, inflammation, and production of a cytokine and/or chemokine. Because prion proteins are generally believed not to evoke an immune response, any prion protein-specific immune response which occurs in association with exposure of an immune cell to a prion protein will generally indicate an immune response to the prion protein.

Also with reference to CpG nucleic acid, an "effective amount to treat the prion disease" shall refer to that amount of CpG nucleic acid that is sufficient to treat a particular prion disease. In one embodiment, an effective amount to treat the prion disease is that amount that is sufficient to slow the development of prion disease, compared to the rate of development of prion disease that would occur without CpG administration according to the instant invention. In one embodiment, an effective amount to treat the prion disease is that amount that is sufficient to prevent the development of prion disease, compared to development of prion disease that would occur without CpG administration according to the instant invention. In one embodiment an effective amount to treat the prion disease is that amount that is sufficient to slow the progression of prion disease, compared to the rate of progression of prion disease that would occur without CpG administration according to the instant invention. In one embodiment an effective amount to treat the prion disease is that amount that is sufficient to stop the progression of prion disease, compared to the progression of prion disease that would occur without CpG administration according to the instant invention. In one embodiment an effective amount to treat the prion disease is that amount that is sufficient to resolve prion disease, compared to the prion disease that would occur without CpG administration according to the instant invention.

Combined with the teachings provided herein, by choosing among the various active compounds and weighing factors such as potency, relative bioavailability, patient body weight, severity of adverse side-effects and preferred mode of administration, an effective prophylactic or therapeutic treatment regimen can be planned which does not cause substantial toxicity and yet is entirely effective to treat the particular subject. The effective amount for any particular application can vary depending on such factors as the disease or condition being treated, the particular immunostimulatory oligonucleotide being administered, the antigen, the size of the subject, or the severity of the disease or condition.

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One of ordinary skill in the art can empirically determine the effective amount of a particular immunostimulatory oligonucleotide and/or other therapeutic agent without necessitating undue experimentation.

Subject doses of the immunostimulatory oligonucleotides for mucosal or local delivery typically range from about 0.1 µg to 10 mg per administration, which depending on the application could be given daily, weekly, or monthly and any other amount of time therebetween. More typically doses range from about 10 µg to 5 mg per administration, and most typically from about 100 µg to 1 mg, with repeated administrations being spaced days or weeks apart. Subject doses of immunostimulatory oligonucleotides for parenteral delivery for the purpose of inducing an antigen-specific immune response, wherein the compounds are delivered with an antigen but not another therapeutic agent are typically 5 to 10,000 times higher than the effective mucosal dose for vaccine adjuvant or immune stimulant applications, and more typically 10 to 1,000 times higher, and most typically 20 to 100 times higher. Doses of the immunostimulatory oligonucleotides for parenteral delivery for the purpose of inducing an innate immune response or for inducing an antigen-specific immune response when the immunostimulatory nucleic acids are administered in combination with other therapeutic agents or in specialized delivery vehicles typically range from about 0.1 µg to 10 mg per administration, which depending on the application could be given daily, weekly, or monthly and any other amount of time therebetween. More typically parenteral doses for these purposes range from about 10 µg to 5 mg per administration, and most typically from about 100 µg to 1 mg, with repeated administrations being spaced days or weeks apart. In some embodiments, however, parenteral doses for these purposes may be used in a range of 5 to 10,000 times higher than the typical doses described above.

For any immunostimulatory oligonucleotide the therapeutically effective amount can be initially determined from animal models. A therapeutically effective dose can also be determined from human data for CpG oligonucleotides which have been tested in humans (human clinical trials have been initiated) and for compounds which are known to exhibit similar pharmacological activities, such as other mucosal adjuvants, e.g., LT and other antigens for vaccination purposes, for mucosal or local administration. Higher doses are required for parenteral administration. The applied dose can be adjusted based on the relative bioavailability and potency of the administered immunostimulatory oligonucleotide.

Adjusting the dose to achieve maximal efficacy based on the methods described herein and

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other methods as are well-known in the art is well within the capabilities of the ordinarily skilled artisan.

The immunostimulatory oligonucleotide can be administered alone or with antigen or other therapeutic agent. In this context, "antigen" refers to any biological molecule capable of eliciting specific immunity. Antigens specifically include peptides (oligopeptides, polypeptides, proteins, and glycosylated derivatives thereof), and polysaccharides. Peptide antigen can be administered preformed or as a polynucleotide encoding the peptide. Also in this context, "other therapeutic agent" includes any suitable composition useful in treating prion disease, including an antibody capable of binding a prion protein. When the immunostimulatory oligonucleotide is administered with antigen or other therapeutic agent, the immunostimulatory oligonucleotide can be administered before, concurrently with, or following administration of the antigen or other therapeutic agent. The immunostimulatory oligonucleotide and the antigen or other therapeutic agent can be formulated together or separately when the immunostimulatory oligonucleotide is administered concurrently with the antigen or other therapeutic agent. When the immunostimulatory oligonucleotide is administered before or following administration of antigen or other therapeutic agent, the immunostimulatory oligonucleotide and the antigen or other therapeutic agent can be administered by the same route of administration or by different routes of administration. In addition, when the immunostimulatory oligonucleotide is administered before or following administration of antigen or other therapeutic agent, the immunostimulatory oligonucleotide and the antigen or other therapeutic agent can be administered to the same site or to different sites.

The immunostimulatory oligonucleotides may be directly administered to the subject or may be administered in conjunction with a nucleic acid delivery complex. A nucleic acid delivery complex shall mean a nucleic acid molecule associated with (e.g., ionically or covalently bound to; or encapsulated within) a targeting means (e.g., a molecule that results in higher affinity binding to target cell (e.g., B cell surfaces and/or increased cellular uptake by target cells). Examples of nucleic acid delivery complexes include nucleic acids associated with a sterol (e.g., cholesterol), a lipid (e.g., a cationic lipid, virosome or liposome), or a target cell specific binding agent (e.g., a ligand recognized by target cell specific receptor). Preferred complexes may be sufficiently stable *in vivo* to prevent significant uncoupling prior to internalization by the target cell. However, the complex can

be cleavable under appropriate conditions within the cell so that the nucleic acid is released in a functional form.

Delivery vehicles or delivery devices for delivering antigen and nucleic acids to surfaces have been described. The immunostimulatory oligonucleotide and/or the antigen and/or other therapeutics may be administered alone (e.g., in saline or buffer) or using any delivery vehicles known in the art. For instance the following delivery vehicles have been described: cochleates (Gould-Fogerite et al., 1994, 1996); emulsomes (Vancott et al., 1998, Lowell et al., 1997); ISCOMs (Mowat et al., 1993, Carlsson et al., 1991, Hu et., 1998, Morein et al., 1999); liposomes (Childers et al., 1999, Michalek et al., 1989, 1992, de Haan 1995a, 1995b); live bacterial vectors (e.g., Salmonella, Escherichia coli, bacillus Calmette-Guérin, Shigella, Lactobacillus) (Hone et al., 1996, Pouwels et al., 1998, Chatfield et al., 1993, Stover et al., 1991, Nugent et al., 1998); live viral vectors (e.g., Vaccinia, adenovirus, Herpes Simplex) (Gallichan et al., 1993, 1995, Moss et al., 1996, Nugent et al., 1998, Flexner et al., 1988, Morrow et al., 1999); microspheres (Gupta et al., 1998, Jones et al., 1996, Maloy et al., 1994, Moore et al., 1995, O'Hagan et al., 1994, Eldridge et al., 1989); nucleic acid vaccines (Fynan et al., 1993, Kuklin et al., 1997, Sasaki et al., 1998, Okada et al., 1997, Ishii et al., 1997); polymers (e.g., carboxymethylcellulose, chitosan) (Hamajima et al., 1998, Jabbal-Gill et al., 1998); polymer rings (Wyatt et al., 1998); proteosomes (Vancott et al., 1998, Lowell et al., 1988, 1996, 1997); sodium fluoride (Hashi et al., 1998); transgenic plants (Tacket et al., 1998, Mason et al., 1998, Haq et al., 1995); virosomes (Gluck et al., 1992, Mengiardi et al., 1995, Cryz et al., 1998); virus-like particles (Jiang et al., 1999, Leibl et al., 1998).

The immunostimulatory oligonucleotides may be administered by any means known to the skilled artisan. Routes of administration include but are not limited to oral, mucosal, parenteral, intravenous, intramuscular, intraperitoneal, intranasal, intratracheal, sublingual, subcutaneous, intradermal, inhalation, ocular, vaginal, and rectal.

The immunostimulatory oligonucleotides are administered in pharmaceutically acceptable solutions, which may routinely contain pharmaceutically acceptable concentrations of salt, buffering agents, preservatives, compatible carriers, adjuvants, and optionally other therapeutic ingredients.

Suitable liquid or solid pharmaceutical preparation forms are, for example, aqueous or saline solutions for inhalation, microencapsulated, encochleated, coated onto microscopic

gold particles, contained in liposomes, nebulized, aerosols, pellets for implantation into the skin, or dried onto a sharp object to be scratched into the skin. The pharmaceutical compositions also include granules, powders, tablets, coated tablets, (micro)capsules, suppositories, syrups, emulsions, suspensions, creams, drops or preparations with protracted release of active compounds, in whose preparation excipients and additives and/or auxiliaries such as disintegrants, binders, coating agents, swelling agents, lubricants, flavorings, sweeteners or solubilizers are customarily used as described above. The pharmaceutical compositions are suitable for use in a variety of drug delivery systems. For a brief review of methods for drug delivery, see Langer R (1990) *Science* 249:1527-33, which is incorporated herein by reference.

The immunostimulatory oligonucleotides and optionally other therapeutics and/or antigens may be administered *per se* (neat) or in the form of a pharmaceutically acceptable salt. When used in medicine the salts should be pharmaceutically acceptable, but non-pharmaceutically acceptable salts may conveniently be used to prepare pharmaceutically acceptable salts thereof. Such salts include, but are not limited to, those prepared from the following acids: hydrochloric, hydrobromic, sulphuric, nitric, phosphoric, maleic, acetic, salicylic, p-toluene sulphonic, tartaric, citric, methane sulphonic, formic, malonic, succinic, naphthalene-2-sulphonic, and benzene sulphonic. Also, such salts can be prepared as alkaline metal or alkaline earth salts, such as sodium, potassium or calcium salts of the carboxylic acid group.

Suitable buffering agents include: acetic acid and a salt (1-2% w/v); citric acid and a salt (1-3% w/v); boric acid and a salt (0.5-2.5% w/v); and phosphoric acid and a salt (0.8-2% w/v). Suitable preservatives include benzalkonium chloride (0.003-0.03% w/v); chlorobutanol (0.3-0.9% w/v); parabens (0.01-0.25% w/v) and thimerosal (0.004-0.02% w/v).

For oral administration, the immunostimulatory oligonucleotides can be formulated readily by combining the active compound(s) with pharmaceutically acceptable carriers well known in the art. Such carriers enable the compounds of the invention to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions and the like, for oral ingestion by a subject to be treated. Pharmaceutical preparations for oral use can be obtained as solid excipient, optionally grinding a resulting mixture, and processing the mixture of granules, after adding suitable auxiliaries, if desired, to obtain tablets or dragee cores. Suitable excipients are, in particular, fillers such as sugars, including lactose, sucrose,

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mannitol, or sorbitol; cellulose preparations such as, for example, maize starch, wheat starch, rice starch, potato starch, gelatin, gum tragacanth, methyl cellulose, hydroxypropylmethyl-cellulose, sodium carboxymethylcellulose, and/or polyvinylpyrrolidone (PVP). If desired, disintegrating agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, or alginic acid or a salt thereof such as sodium alginate. Optionally the oral formulations may also be formulated in saline or buffers for neutralizing internal acid conditions or may be administered without any carriers.

Dragee cores are provided with suitable coatings. For this purpose, concentrated sugar solutions may be used, which may optionally contain gum arabic, talc, polyvinyl pyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for identification or to characterize different combinations of active compound doses.

Pharmaceutical preparations which can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a plasticizer, such as glycerol or sorbitol. The push-fit capsules can contain the active ingredients in admixture with filler such as lactose, binders such as starches, and/or lubricants such as talc or magnesium stearate and, optionally, stabilizers. In soft capsules, the active compounds may be dissolved or suspended in suitable liquids, such as fatty oils, liquid paraffin, or liquid polyethylene glycols. In addition, stabilizers may be added. Microspheres formulated for oral administration may also be used. Such microspheres have been well defined in the art. All formulations for oral administration should be in dosages suitable for such administration.

For buccal administration, the compositions may take the form of tablets or lozenges formulated in conventional manner.

For administration by inhalation, the compounds for use according to the present invention may be conveniently delivered in the form of an aerosol spray presentation from pressurized packs or a nebulizer, with the use of a suitable propellant, e.g., dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide or other suitable gas. In the case of a pressurized aerosol the dosage unit may be determined by providing a valve to deliver a metered amount. Capsules and cartridges of e.g., gelatin for use in an inhaler or insufflator may be formulated containing a powder mix of the compound and a suitable powder base such as lactose or starch.

The immunostimulatory oligonucleotides, when it is desirable to deliver them systemically, may be formulated for parenteral administration by injection, e.g., by bolus injection or continuous infusion. Formulations for injection may be presented in unit dosage form, e.g., in ampoules or in multi-dose containers, with an added preservative. The compositions may take such forms as suspensions, solutions or emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilizing and/or dispersing agents.

Pharmaceutical formulations for parenteral administration include aqueous solutions of the active compounds in water-soluble form. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes. Aqueous injection suspensions may contain substances which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Optionally, the suspension may also contain suitable stabilizers or agents which increase the solubility of the compounds to allow for the preparation of highly concentrated solutions.

Alternatively, the active compounds may be in powder form for constitution with a suitable vehicle, e.g., sterile pyrogen-free water, before use.

The immunostimulatory oligonucleotides may also be formulated in rectal or vaginal compositions such as suppositories or retention enemas, e.g., containing conventional suppository bases such as cocoa butter or other glycerides.

In addition to the formulations described previously, the immunostimulatory oligonucleotides may also be formulated as a depot preparation. Such long acting formulations may be formulated with suitable polymeric or hydrophobic materials (for example as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives, for example, as a sparingly soluble salt.

The pharmaceutical compositions also may comprise suitable solid or gel phase carriers or excipients. Examples of such carriers or excipients include but are not limited to calcium carbonate, calcium phosphate, various sugars, starches, cellulose derivatives, gelatin, and polymers such as polyethylene glycols.

The term "pharmaceutically-acceptable carrier" means one or more compatible solid or liquid filler, diluents or encapsulating substances which are suitable for administration to a

human or other vertebrate animal. The term "carrier" denotes an organic or inorganic ingredient, natural or synthetic, with which the active ingredient is combined to facilitate the application. The components of the pharmaceutical compositions also are capable of being commingled with the compounds of the present invention, and with each other, in a manner such that there is no interaction which would substantially impair the desired pharmaceutical efficiency.

The immunostimulatory oligonucleotides useful in the invention may be delivered in mixtures with additional adjuvant(s), other therapeutics, or antigen(s). A mixture may consist of several adjuvants in addition to the immunostimulatory oligonucleotide or several antigens or other therapeutics.

The particular mode selected will depend, of course, upon the particular adjuvants or antigen selected, the particular condition being treated and the dosage required for therapeutic efficacy. The methods of this invention, generally speaking, may be practiced using any mode of administration that is medically acceptable, meaning any mode that produces effective levels of an immune response without causing clinically unacceptable adverse effects. Preferred modes of administration are discussed above.

The immunostimulatory oligonucleotides may conveniently be presented in unit dosage form and may be prepared by any of the methods well known in the art of pharmacy. All methods include the step of bringing the immunostimulatory oligonucleotides into association with a carrier which constitutes one or more accessory ingredients. In general, the compositions are prepared by uniformly and intimately bringing the compounds into association with a liquid carrier, a finely divided solid carrier, or both, and then, if necessary, shaping the product. Liquid dose units are vials or ampoules. Solid dose units are tablets, capsules and suppositories. For treatment of a patient, depending on activity of the compound, manner of administration, purpose of the immunization (i.e., prophylactic or therapeutic), nature and severity of the disorder, age and body weight of the patient, different doses may be necessary. The administration of a given dose can be carried out both by single administration in the form of an individual dose unit or else several smaller dose units. Multiple administration of doses at specific intervals of weeks or months apart is usual for boosting the antigen-specific responses.

Other delivery systems can include time-release, delayed release or sustained release delivery systems. Such systems can avoid repeated administrations of the compounds,

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increasing convenience to the subject and the physician. Many types of release delivery systems are available and known to those of ordinary skill in the art. They include polymer base systems such as poly(lactide-glycolide), copolyoxalates, polycaprolactones, polyesteramides, polyorthoesters, polyhydroxybutyric acid, and polyanhydrides. Microcapsules of the foregoing polymers containing drugs are described in, for example, U.S. Patent 5,075,109. Delivery systems also include non-polymer systems that are: lipids including sterols such as cholesterol, cholesterol esters and fatty acids or neutral fats such as mono-di-and tri-glycerides; hydrogel release systems; sylastic systems; peptide based systems; wax coatings; compressed tablets using conventional binders and excipients; partially fused implants; and the like. Specific examples include, but are not limited to: (a) erosional systems in which an agent of the invention is contained in a form within a matrix such as those described in U.S. Patent Nos. 4,452,775, 4,675,189, and 5,736,152, and (b) diffusional systems in which an active component permeates at a controlled rate from a polymer such as described in U.S. Patent Nos. 3,854,480, 5,133,974 and 5,407,686. In addition, pump-based hardware delivery systems can be used, some of which are adapted for implantation.

Examples

Example 1. CpG-ODN Protects Mice from Scrapie Prion.

Groups of 8 mice each were inoculated intraperitoneally with 100 μl of 10% brain homogenates from mice terminally ill with RML scrapie prion strain corresponding to an infectious challenge of approximately 10⁴ LD₅₀. The mice received 0.15 pmol (30 μl of 5 nM solution) CpG-ODN (oligonucleotide 1826, 5'- TCCATGACGTTCCTGACGTT -3', SEQ ID NO:18), which has been shown to be a strong inducer of innate immunity and to confer sterile immunity against certain infectious diseases. Sparwasser T et al. (2000) *Eur J Immunol* 30:3591-7. CpG-ODN was administered intraperitoneally at the time of inoculation (0h) with scrapie prions as well as 4 times at intervals of 24h (4 x q 24h); a second group received 0.15 pmol (30 μl of 5 nM solution) CpG-ODN 7h after infection (7h) and 4 times at intervals of 24h (4 x q 24h); a third group received CpG-ODN at 7h after infection and subsequently 20 times at intervals of 24h (20 x q 24h). Controls matched for age and sex were given saline instead of CpG-ODN at the identical time intervals. Additional control experiments were also performed in uninfected mice using saline and brain homogenates of

uninfected mice. All animals were observed and scored daily for clinical signs of disease. Scrapie in mice is characterized by ataxia of gait, tremor, difficulty righting from a supine position, and tail rigidity. Occurrence of two of these four symptoms was used as the end point criterion for establishing a clinical diagnosis of scrapie. Western blots of brain homogenates were performed to confirm the diagnosis. All results were analyzed using the Student's t test (Table 1).

Table 1. Mean incubation time in C57BL/6 mice after inoculation with scrapie strain RML and treatment with CpG-ODN.

Group	Inoculation	Treatment	Regimen	Time to Terminal Disease (d) ± SD	Attack Rate
1	RML	CpG 1826	0h and 4 x q 24h	253 ± 4	8/8
2	RML	CpG 1826	7h and 4 x q 24h	250 ± 6	8/8
3	RML	CpG 1826	7h and 20 x q 24h	> 330 no disease	0/8
4	RML	Saline	0h and 4 x q 24h	183 ± 7	8/8
5	RML	Saline	7h and 4 x q 24h	181 ± 3	8/8
6	· RML =	⊹Saline ~	:7h and 20 x q 24h	181 ± 3	8/8
7	'-Saline €	CpG 1826	-0h and 4 x q 24h	No disease	0/8
8	Saline	CpG 1826	7h and 4 x q 24h	No disease	0/8
9	Saline	CpG 1826	7h and 20 x q 24h	No disease	0/8
10	Brain homogenate of uninfected mouse	CpG 1826	0h and 4 x q 24h	No disease	0/8
11	Brain homogenate of uninfected mouse	CpG 1826	7h and 4 x q 24h	No disease	0/8
12	Brain homogenate of uninfected mouse	CpG 1826	7h and 20 x q 24h	No disease	0/8

Mice infected with the RML strain which received CpG-ODN at the time of inoculation and 7h post-infection as well as 4 times at intervals of 24h showed a dramatic prolongation of survival time compared to control mice with an increase in survival time of 38% in both cases. These differences were highly significant (p<0.0001). The application of CpG-ODN 7h post-inoculation and 20 times at 24h intervals led to disease-free intervals of more than 330 days. All control groups which were not inoculated with the RML strain remained disease-free, and no harmful effect of CpG-ODN application was observed.

These results showed that the application of CpG-ODN at the time of infection and 7h after infection led to a dramatic prolongation of survival time. This effect can be amplified when CpG-ODN are given for a longer period of time. The application of CpG-ODN for 20 times at 24h intervals results in a disease-free interval of >330 days, which indicates the great potential of CpG-ODN for post-exposure prophylaxis of people with exposure to infection. The mechanism of disease prevention remains to be determined, but it seems that the most likely explanation for this effect is a stimulation of TLR-expressing cells of the innate immune system, e.g., macrophages, monocytes, and especially dendritic cells. Sparwasser T et al. (2000) Eur J Immunol 30:3591-7. CpG-ODN has been known to induce resistance against other infectious diseases. Zimmermann S et al. (1998) J Immunol 160:3627-30. The induction of extreme prolongation of the incubation time or even resistance to prion disease was a surprising finding in the context of a completely different infectious agent.

The findings presented here show that administration of CpG-ODN prolongs the incubation time by 38% and may have the potential to prevent infection after repeated administration, even when high doses of infectivity are administered intraperitoneally. It may therefore be possible to prevent disease after inadvertent intraperitoneally. It may lower infectious doses administered peripherally.

Example 2. Effect of Timing Between Exposure to Prion and Administration of CpG-ODN.

Mice are injected with RML scrapie prion or control and treated with CpG-ODN or control essentially as in Example 1, except that the interval between injection with RML scrapie prion or control and administration of CpG-ODN or control is varied. In some groups administration of CpG-ODN is delayed as much as a month following injection with RML scrapie prion or control. In some groups administration of CpG-ODN precedes injection with RML scrapie prion or control. In some groups the number and schedule of repeated administrations of CpG-ODN is varied from Example 1. Results, measured as in Example 1, show that CpG-ODN is effective even when administered more than 7h after injection with RML scrapie prion.

Example 3. Mice Protected from Scrapie Prion by CpG-ODN Develop an Immune Response to Prion.

Mice are injected with RML scrapie prion or control and treated with CpG-ODN or control essentially as in Example 1 or Example 2. At various time points following injection with RML scrapie prion or control and treatment with CpG-ODN or control, tissue or blood samples are obtained and analyzed for prion-specific and prion-nonspecific immune response. Presence of an immune response is determined by suitable method or measurement including, without limitation, antibody titer, enzyme-linked immunosorbent assay (ELISA), flow cytometry, cell proliferation assay, cytotoxicity assay, polymerase chain reaction (PCR) and reverse transcriptase-polymerase chain reaction (RT-PCR), Western immunoblot, Northern blot, and Southern blot. General methods for these types of measurements are standard and are suitably adapted to the specific antigen or stimulus being assayed. For example, ELISA is used to measure production of various secreted products, including antibodies, cytokines and chemokines. Cytokines and chemokines in this example include interleukin (IL)-4, IL-10, IL-6, IL-12, IL-18, interferon (IFN)-α, IFN-β, IFN-γ, tumor necrosis factor (TNF), and IP-10. Flow cytometry is used to measure cell surface and intracellular proteins, including markers associated with immune cell activation. Markers associated with immune cell activation can vary with cell type but include cluster of differentiation (CD) markers such as CD86; major histocompatibility complex (MHC), inducible cytokine receptors, and certain costimulatory molecules. Results show that CpG-ODN induces an immune response to prion protein.

Example 4. Selection of CpG-ODN.

Mice are injected with RML scrapie prion or control and treated with CpG-ODN or control essentially as in Example 1 or Example 2. Various CpG-ODN are compared against ODN 1826 for their effectiveness. Results, measured as in Example 1, show that protection is related to the use of species-optimized CpG-ODN.

Example 5. Use of CpG-ODN in Alzheimer's Disease Model.

Mice genetically susceptible to developing Alzheimer's-like disease are administered CpG nucleic acid alone or CpG nucleic acid plus antigen (e.g., amyloid precursor protein or Aβ), either prior to or following onset of Alzheimer's-like disease. Similar mice are administered appropriate control treatment. Animals are monitored for behavioral and histologic evidence of Alzheimer's-like disease.

The foregoing written specification is considered to be sufficient to enable one skilled in the art to practice the invention. The present invention is not to be limited in scope by examples provided, since the examples are intended as a single illustration of one aspect of the invention and other functionally equivalent embodiments are within the scope of the invention. Various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and fall within the scope of the appended claims. The advantages and objects of the invention are not necessarily encompassed by each embodiment of the invention.

All references, patents and patent publications that are recited in this application are incorporated in their entirety herein by reference.

We claim:

Claims

- A method for treating a prion disease in a subject, comprising:
 administering to a subject having or at risk of developing a prion disease a CpG
 nucleic acid in an effective amount to treat the prion disease.
- 2. The method according to claim 1, wherein the administering follows exposure of the subject to a prion protein that is associated with a prion disease.
- 3. The method according to claim 1, wherein the prion disease is a transmissible spongiform encephalopathy (TSE).
- 4. The method according to claim 1, wherein the prion disease is scrapie.
- 5. The method according to claim 1, wherein the prion disease is bovine spongiform encephalopathy (BSE).
- 6. The method according to claim 1, wherein the prion disease is variant Creutzfeldt-Jakob disease (vCJD).
- 7. The method according to claim 1, wherein the prion disease is iatrogenic Creutzfeldt-Jakob disease (iCJD).
- 8. The method according to claim 1, wherein the subject is a human.
- 9. A method for inducing an immune response to a prion protein, comprising: contacting an antigen-presenting cell (APC) with a prion protein; and contacting the APC with a CpG nucleic acid in an effective amount to induce an immune response to the prion protein.
- 10. The method according to claim 9, wherein the immune response is in vivo.

- 11. The method according to claim 9, wherein the APC is selected from the group consisting of: a B cell, a dendritic cell, a macrophage, and a monocyte.
- 12. The method according to claim 9, wherein the APC is a dendritic cell.
- 13. The method according to claim 9, wherein the APC expresses a Toll-like receptor (TLR) that signals in response to the CpG nucleic acid.
- 14. The method according to claim 13, wherein the TLR is TLR9.
- 15. The method according to claim 9, wherein the prion protein is prion protein:scrapie form (PrPSc).
- 16. The method according to claim 9, wherein the prion protein is a fragment of PrP^{Sc} lacking at least the amino terminus of full-length PrP^{Sc}.
- 17. The method according to claim 9, wherein the prion protein is a derivative of PrP^{Sc} or a derivative of a fragment of PrP^{Sc} lacking at least the amino terminus of full-length PrP^{Sc}.
- 18. The method according to claim 9, wherein the CpG nucleic acid is a Class B CpG nucleic acid.
- The method according to claim 9, wherein the CpG nucleic acid is a Class A CpG nucleic acid.
- 20. The method according to claim 9, wherein the CpG nucleic acid is a Class C CpG nucleic acid.
- 21. The method according to claim 9, wherein the CpG nucleic acid is optimized for use in a species of the subject.

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- Asn Arg Tyr Pro Pro Gln Gly Gly Gly Gly Trp Gly Gln Pro His Gly 50 55 60
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- Thr Glu Thr Asp Ile Lys Met Met Glu Arg Val Val Glu Gln Met Cys 210 215 220
- Val Thr Gln Tyr Gln Lys Glu Ser Gln Ala Tyr Tyr Asp Gln Gly Ala 225 230 235 240

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Gly Asn Arg Tyr Pro Pro Gln Gly Gly Gly Gly Trp Gly Gln Pro His 50 55 60

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Met Ser Arg Pro Leu Ile His Phe Gly Ser Asp Tyr Glu Asp Arg Tyr 145 - 150 - 155 160

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Cys Ile Thr Gln Tyr Gln Arg Glu Ser Gln Ala Tyr Tyr Gln Arg Gly 225 230 235 240

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